

THE SYNTHESIS AND
ANTIBACTERIAL ACTIVITY OF SOME
 β -LACTAMASE STABLE
6 α -(HYDROXYMETHYL)PENICILLINS

RONALD A. DIXON, ROBERT A. EDMONDSON,
KENNETH D. HARDY and PETER H. MILNER

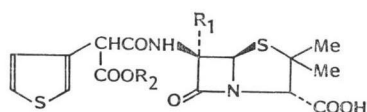
Beecham Pharmaceuticals, Research Division,
Brockham Park, Betchworth, Surrey,
RH3 7AJ, U. K.

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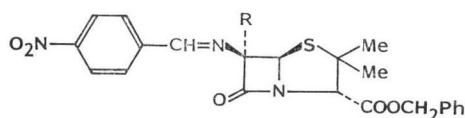
The emergence of bacterial resistance to antibiotics has long been a problem in the clinic. In the case of penicillins and cephalosporins, resistance is often due to the action of β -lactamase, enzymes which inactivate the antibiotic by hydrolysis of the β -lactam ring¹. Certain penicillins² are available which display activity against β -lactamase producing Gram-positive organisms, and some hitherto resistant Gram-negative bacteria can be treated by a number of cephalosporins. However, despite attempts by several groups³⁻⁵, it

is only recently with the development of temocillin (BRL 17421)⁶ (1) that a β -lactamase stable penicillin with good Gram-negative activity has been produced. We now wish to report the preparation and antibacterial activity of some 6 α -(hydroxymethyl)penicillins¹⁰, which are further examples of penicillins showing activity against a variety of β -lactamase producing Gram-negative bacteria.

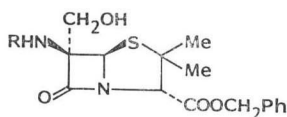
Introduction of the hydroxymethyl group was accomplished by a modification of the route employed by Merck workers^{4,11}. Condensation of *p*-nitrobenzaldehyde with benzyl 6 β -amino-penicillanate in the presence of 4 Å molecular sieves, afforded the Schiff's base 4. Treatment of 4 in *N,N*-dimethylformamide at 0°C with anhydrous potassium carbonate followed by gaseous formaldehyde gave the 6 α -(hydroxymethyl) analogue (5), which with toluene-*p*-sulfonic acid monohydrate in ethyl acetate afforded the protected nucleus (6) as its toluene-*p*-sulfonic acid salt. The free amine was liberated from this salt by reaction with aqueous sodium hydrogen carbonate, and could be acylated with the appro-



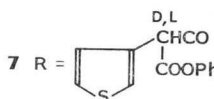
- 1 $R_1 = \text{OCH}_3$, $R_2 = \text{H}$
2 $R_1 = \text{CH}_2\text{OH}$, $R_2 = \text{Ph}$
3 $R_1 = \text{CH}_2\text{OH}$, $R_2 = \text{H}$



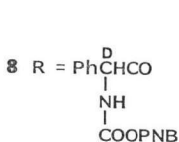
- 4 $R = \text{H}$
5 $R = \text{CH}_2\text{OH}$



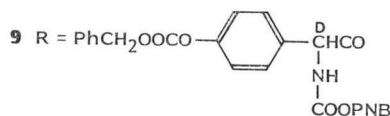
6 $R = \text{H}$



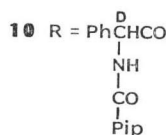
7 $R =$



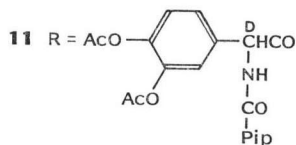
8 $R = \text{Ph}$



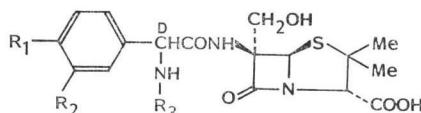
9 $R = \text{PhCH}_2\text{OOCO}$



10 $R = \text{Ph}$



11 $R = \text{AcO}$



12 $R_1 = R_2 = R_3 = \text{H}$

13 $R_1 = \text{OH}$, $R_2 = R_3 = \text{H}$

14 $R_1 = R_2 = \text{H}$, $R_3 = \text{CO-Pip}$

15 $R_1 = \text{OH}$, $R_2 = \text{H}$, $R_3 = \text{CO-Pip}$

16 $R_1 = R_2 = \text{OAc}$, $R_3 = \text{CO-Pip}$

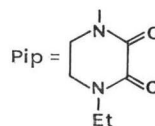
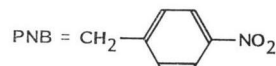


Table 1. The relative activities *in vitro* of compounds **14**~**16** and piperacillin.

Organism	MIC ($\mu\text{g/ml}$) ^c			
	14	15	16	Piperacillin
<i>Escherichia coli</i> ESS	0.1	0.1	<0.06	<0.06
<i>E. coli</i> JT 4 ^a	100	100	1.0	>128
<i>E. coli</i> JT425 ^b	50	50	8.0	16
<i>E. coli</i> NCTC 10418	25	5.0	0.12	0.5
<i>Pseudomonas aeruginosa</i> NCTC 10662	>100	>100	4.0	4.0
<i>P. aeruginosa</i> Dalgleish ^a	>100	>100	8.0	>128
<i>Serratia marcescens</i> US 32	50	5.0	2.0	1.0
<i>Klebsiella aerogenes</i> A	5.0	2.5	0.12	2.0
<i>Enterobacter cloacae</i> N1	25	5.0	4.0	1.0
<i>Proteus mirabilis</i> C977	10	2.5	4.0	0.5
<i>P. mirabilis</i> 889 ^b	5.0	2.5	4.0	>128
<i>P. rettgeri</i>	50	50	8.0	0.5
<i>Staphylococcus aureus</i> Oxford	>100	>100	>128	0.5
<i>S. aureus</i> Russell ^a	>100	>100	>128	>128
<i>Streptococcus pyogenes</i> CN10	25	25	16	0.12

^a β -Lactamase producing strain (plasmid-mediated).

^b β -Lactamase producing strain (non-plasmid-mediated).

^c Determined by serial dilution in nutrient agar containing 5% defibrinated horse blood, inoculum 0.001 ml of an undiluted overnight broth culture (approximately 10^8 cfu).

Table 2. Stability of **14** and **15** to cell-free β -lactamase preparations from Gram-negative and Gram-positive bacteria.

Enzyme preparation	Class of β -lactamase ¹²⁾	Stability ^a		
		14	15	Piperacillin
<i>Escherichia coli</i> R ⁺ TEM	Plasmid mediated penicillinase III a	NDH ^b	NDH	5.3×10^3
<i>E. coli</i> JT414	Chromosomally mediated cephalosporinase Ib	2	NDH	20
<i>Enterobacter cloacae</i> 10005	Chromosomally mediated cephalosporinase Ia	3	6	NDH
<i>Proteus mirabilis</i> 889	Chromosomally mediated penicillinase II	NDH	NDH	5.3×10^3
<i>Klebsiella pneumoniae</i> A	Chromosomally mediated penicillinase IV	NDH	NDH	NDH
<i>Staphylococcus aureus</i> MB9	Gram-positive penicillinase	NDH	NDH	9.6×10^2

^a Figures are rates of hydrolysis ($\mu\text{M}/\text{hour}$) for 1 cell enzyme unit, (the concentration of β -lactamase produced after 7 hours growth in nutrient broth No. 2 at 37°C).

Reaction mixtures containing 0.1 mM substrate were made up in 0.05 M phosphate buffer (pH 7).

Samples removed in duplicate at 0, 10, 20 and 40 minutes, and the residual substrate determined by a tape bioassay method¹³⁾ using *E. coli* ESS.

^b No detectable hydrolysis at 5 cell enzyme units.

priate side-chain acids to afford a series of 6 β -acylamino-6 α -(hydroxymethyl)penicillanates (**7**~**11**). Hydrogenolysis of **7** over 10% palladium on charcoal catalyst afforded the half-ester (**2**), which on hydrolysis with aqueous sodium tetraborate yielded 6 α -(hydroxymethyl)ticarcillin (**3**). Deprotection (10% Pd/C; H₂) of **8** and

9 gave the 6 α -(hydroxymethyl)ampicillin (**12**) and amoxicillin (**13**) derivatives respectively, which could be further acylated to afford the piperacillin derivatives **14** and **15**. Preparation of **14** could also be effected by hydrogenolysis of the benzyl ester (**10**), and in a similar manner, deprotection of **11** gave the 3,4-diacetoxyphenyl-

penicillin (16).

The stereochemistry of the hydroxymethylation reaction has been assigned by the Merck group⁴⁾ as occurring from the α -face of the Schiff's base (4). We have confirmed this by demonstrating a positive nuclear Overhauser effect ($16 \pm 9\%$) between the 5α -proton and the 6-methylene group in the 6α -(hydroxymethyl)piperacillin ester (10).

Several of the compounds prepared including an α -carboxy analogue (3) and α -amino derivatives 12 and 13 were essentially inactive. However compounds 14~16, which are structurally related to piperacillin, inhibited many clinically derived Gram-negative bacteria (Table 1). In particular the 3,4-diacetoxyphenyl derivative (16) showed good activity against β -lactamase producing strains of *Escherichia coli*, *Pseudomonas aeruginosa* and *Proteus mirabilis*. None of these compounds displayed significant activity against Gram-positive organisms.

The stabilities of compounds 14 and 15 to isolated bacterial β -lactamases, compared with piperacillin, are shown in Table 2. Neither of the 6α -(hydroxymethyl)penicillins were significantly hydrolysed, whereas piperacillin was inactivated by enzymes isolated from *E. coli*, *P. mirabilis* and *Staphylococcus aureus*. Thus, incorporation of the 6α -(hydroxymethyl) group into the penicillin nucleus, like the 6α -methoxy substituent in temocillin, confers improved stability to bacterial β -lactamases.

Acknowledgments

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